Search notes

Search Results -

Terms	Documents
(demembranated or membrane adj disrupted) and sperm	10

Database: US Patents Full-Text Database

Refine Search: (demembranated or membrane adj disrupted) and sperm

Search History

DB Name	Query	Hit Count	Set Name
USPT	(demembranated or membrane adj disrupted) and sperm	10	L8
USPT	l5 and transfer	2514	L7
USPT	gene adj transger and sperm	0	L6
USPT	gene and sperm	4398	L5
USPT	transfer and sperm	3008	L4
USPT	transgenic and sperm adj head	5	L3
USPT	transgenic and sperm	1147	L2
USPT	transgenic	5046	L1

L23 ANSWER 1 OF 16 CAPLUS COPYRIGHT 2000 ACS
ACCESSION NUMBER: 2000:133382 CAPLUS
TITLE: Method of performing transgenesis
INVENTOR(S): Perry, Anthony C. F.; Wakayama, Teruhiko
PATENT ASSIGNEE(S): USA
SOURCE: PCT Int. Appl., 45 pp.
CODEN: PIXXD2
DOCUMENT TYPE: Patent
LANGUAGE: English
FAMILY ACC. NUM. COUNT: 2
PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 2000008924	A1	20000224	WO 1999-US18429	19990811

W: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM

RW: GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG

PRIORITY APPLN. INFO.:		
US 1998-96078	19980811	
US 1999-134251	19990513	

AB The invention provides a method for generating transgenic animals and cells by the coinsertion of nucleic acid and a nucleus into an unfertilized oocyte. Preferably, the coinsertion is by **microinjection** and more preferably by piezo-electrically actuated **microinjection**. Transgene (tg) expressing embryos are here produced following coinjection of unfertilized mouse oocytes with **sperm heads** and exogenous DNA encoding either a green fluorescent protein (GFP) or .beta.-galactosidase reporter. The microinjected oocyte may be allowed to develop into differentiated cells or stem cells; into an embryo in vitro prior to transfer into a host surrogate mother; or it may be transferred directly into a host surrogate mother. Embryonic development can occur to term, such that the offspring possess transgenic modifications that may alter their characteristics (phenotype) and are, in turn, transmitted to their offspring.

L7 ANSWER 4 OF 10 MEDLINE DUPLICATE 1
ACCESSION NUMBER: 1998132258 MEDLINE
DOCUMENT NUMBER: 98132258
TITLE: Behavior of transgenic mouse spermatozoa with galline protamine.
AUTHOR: Maleszewski M; Kuretake S; Evenson D; Yanagimachi H; Bjordahl J; Yanagimachi R
CORPORATE SOURCE: Department of Anatomy and Reproductive Biology, University of Hawaii, Honolulu 96822, USA.
CONTRACT NUMBER: HD-03402 (NICHD)
SOURCE: BIOLOGY OF REPRODUCTION, (1998 Jan) 58 (1) 8-14. Journal code: A3W. ISSN: 0006-3363.
PUB. COUNTRY: United States
Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Priority Journals
ENTRY MONTH: 199805
ENTRY WEEK: 19980502

AB General morphology, physical and chemical stability of nuclei, and postfertilization behavior of spermatozoa from transgenic mice [TgN (Prml Gal) 223 Bri] containing nuclear **avian** protamine (galline) were compared to those in the spermatozoa of wild-type (Wild) mice. Galline to protamine I ratios in spermatozoal nuclei of transgenic mice, strains 3175 (T75) and 3177 (T77), were 1.94 and 5.62, respectively. Live T75 and T77 spermatozoa were indistinguishable in their gross morphology from Wild spermatozoa. However, unlike Wild and T75 spermatozoa, T77 spermatozoa were vulnerable to mechanical handling, as about 40% of heads and tails were separated after gentle pipetting in suspension. Motility of T77 spermatozoa was markedly inferior to that of T75 and Wild. Chromatin heterogeneity and instability of transgenic spermatozoal nuclei were evident by transmission electron microscopy, staining reaction to Giemsa, and, as apparent by both light microscopy and flow cytometry, reaction to SDS detergent. Wild and T75 spermatozoa fertilized 90% and 60% of zona-intact **oocytes** in vitro, respectively. T77 spermatozoa completely failed to fertilize and bound to zona surfaces very weakly, and none of them inserted their heads into the zona. Although inefficiently, T77 spermatozoa could fertilize zona-free **oocytes** in vitro, indicating some ability to undergo capacitation and spontaneous acrosome reaction in vitro. After microsurgical injection into **oocytes**, the rate of nuclear decondensation was the greatest in rooster spermatozoa, followed by T77, T75, and Wild spermatozoa.

L9 ANSWER 6 OF 12 EMBASE COPYRIGHT 2000 ELSEVIER SCI. B.V.
ACCESSION NUMBER: 96168878 EMBASE
DOCUMENT NUMBER: 1996168878
TITLE: **Culture** of naked quail (Coturnix coturnix japonica) ova in vitro for **avian** transgenesis: **Culture** from the single-cell stage to hatching with pH-adjusted chicken thick albumen.
AUTHOR: Ono T.; Murakami T.; Tanabe Y.; Mizutani M.; Mochii M.; Eguchi G.
CORPORATE SOURCE: Laboratory of Developmental Biology, Faculty of Agriculture, Shinshu University,Ina 399-45, Japan
SOURCE: Comparative Biochemistry and Physiology - A Physiology, (1996) 113/3 (287-292).
ISSN: 0300-9629 CODEN: CBPAB5
COUNTRY: United States
DOCUMENT TYPE: Journal; Article
FILE SEGMENT: 021 Developmental Biology and Teratology
029 Clinical Biochemistry
LANGUAGE: English
SUMMARY LANGUAGE: English

AB We first examined the pH change of the albumen of quail (Coturnix coturnix japonica) eggs before and after they were laid, as well as that of laid eggs. The pH rose rapidly after laying and continued to increase gradually in storage. Incubation at 37.5.degree.C accelerated the increase in the pH of infertile eggs, while that of fertile eggs remained low during incubation. Referring to these results, we obtained a protocol for producing quail hatchlings by **culture** in vitro from naked ova. The naked ovum was filled with chicken (Gallus domesticus) thick albumen, the pH of which had been adjusted to 7.2-7.4. The ovum was cultured at 41.5.degree.C in 20% CO2 in air for the first 24 h. Then the embryo was moved to a surrogate quail egg shell that had been filled with non-pH-adjusted chicken thin albumen and cultured for a further 48 h in 100% air. The embryo was transferred again to a surrogate chicken egg shell and cultured under the same conditions until hatching. The **culture** yielded quail chicks with a hatchability of 19.4%. The method proposed here should be applicable to the production of transgenic birds.

L7 ANSWER 3 OF 10 BIOSIS COPYRIGHT 2000 BIOSIS
ACCESSION NUMBER: 1999:289854 BIOSIS
DOCUMENT NUMBER: PREV199900289854
TITLE: **Fertilization** and early **avian** development.
AUTHOR(S): Stepinska, Urszula (1)
CORPORATE SOURCE: (1) Jastrzebiec, 05-551, Mrokow Poland
SOURCE: Postepy Biologii Komorki, (1999) Vol. 26, No. SUPPL. 12, pp. 73-78.
ISSN: 0324-833X.
DOCUMENT TYPE: Article
LANGUAGE: Polish
SUMMARY LANGUAGE: English; Polish

AB The early developmental stages of the bird are rather poorly understood and are the subject of endless discussion. The reason for this has been the difficulty in obtaining the experimental material - the **fertilization** and early embryogenesis (cleavage and area pellucida formation) take place in the hen's oviduct, besides, only 1 oocyte is ovulated every 24 hr. Birds exibit physiological polyspermy, i. e. many sperms enter the egg, however only one of them participates in the formation of zygote nucleus, whereas the rest of them degenerate at early cleavage stages. This could suggest the presence of some kind of late block to polyspermy in the cytoplasm of **avian** egg. However, the factors participating in the block are not known. It is suggested that DNase activity present in the germinal discs of quail preovulatory **oocytes**, might be responsible for degradation of supernumerary sperm DNA in the early **avian** embryo.

21 ANSWER 3 OF 7 BIOSIS COPYRIGHT 2000 BIOSIS
ACCESSION NUMBER: 1998:343776 BIOSIS
DOCUMENT NUMBER: PREV199800343776
TITLE: Morphology of a sterile, tetraploid, hybrid whiptail lizard
(Squamata: Teiidae: Cnemidophorus.
AUTHOR(S): Hardy, Laurence M. (1); Cole, Charles J.
CORPORATE SOURCE: (1) Dep. Herpetol., American Museum Natural History {a} Dep. Herpetol., American Museum Natural History USA
SOURCE: American Museum Novitates, (June 10, 1998) Vol. 0, No. 3228, pp. 1-16.
ISSN: 0003-0082.
DOCUMENT TYPE: Article
LANGUAGE: English

AB Experimental hybridization with whiptail lizards has been conducted in order to improve understanding of the evolution of parthenogenesis in vertebrates and the effects of horizontal **gene transfer** in Cnemidophorus, the systematics of which has been confused owing to the reticulate phylogeny within the genus. Here we describe the external morphology and reproductive tissue histology of a sterile tetraploid hybrid between C. sonorae (triploid, unisexual) X C. tigris (diploid, bisexual), and compare her to her parents and siblings that developed from unfertilized eggs (normally cloned C. sonorae). This may help to identify F1 hybrids that are found in nature and may help to determine whether they are sterile without conducting extensive laboratory breeding programs. Considering that the maternal parent (C. sonorae) represented a clone that was of hybrid origin itself, the four genomes in the tetraploid hybrid historically were derived from three hybridization events among three bisexual species of Cnemidophorus, probably as follows: ((inornatus female X burti male) X burti male) X tigris male. The tetraploid inherited 100% of its mother's genes and morphologically was very similar to her and her cloned offspring, particularly in scalation. Nevertheless, it was slightly larger than its siblings at hatching, grew faster than its siblings, attained a larger size, and, beginning at an age of six months, developed dorsal spots reflecting paternal traits in its color pattern. However, if this lizard had been found in nature, without any knowledge of its life history and in the absence of genetic data, it could easily have been misidentified as Cnemidophorits exsanguis, which it resembled more closely than its parental species. Although she reached adult size and lived for more than two years beyond the age at which her cloned siblings produced offspring (nine months), the tetraploid never reproduced. Her ovaries were abnormally small, had poorly defined follicular epithelium with little vascularization, and had either empty or fluid-filled follicles devoid of oocytes. She also had numerous abnormally large mesonephric tubules and few or no cilia in the median oviduct. These traits should be examined in other specimens hypothesized to be sterile F1 hybrid females.